

Diarrhoea Dialogue



ISSUE No. 11

NOVEMBER
1982

Control programmes: the key role of laboratories

Oral rehydration therapy (ORT), properly given, is always the first-line treatment for diarrhoea. Nevertheless, there is only one effective way to control this problem. The causes must be identified and the sources of infection traced and treated within communities.

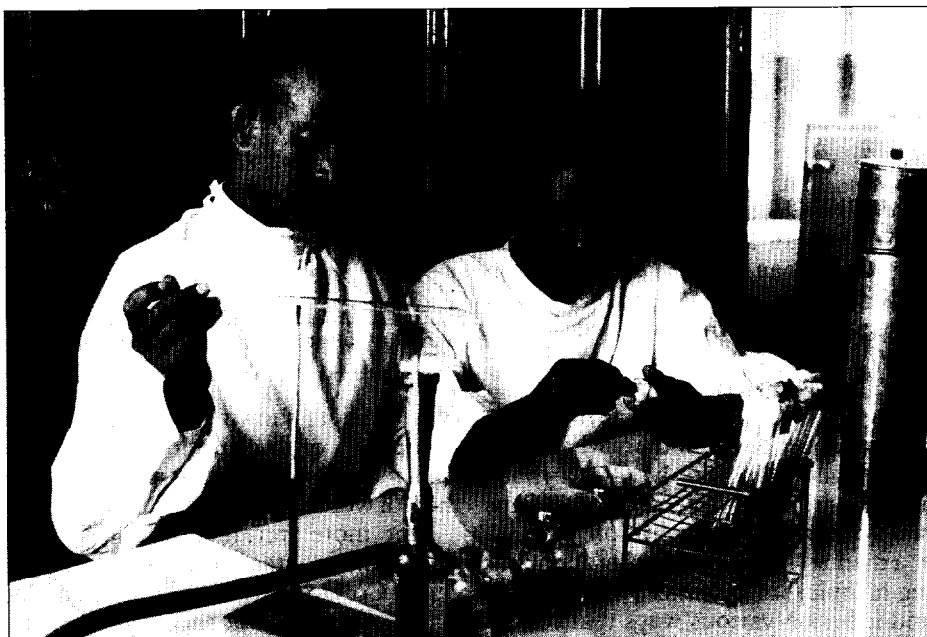
Advances in techniques

Until recently control programmes have been handicapped because, up to ten years ago, the exact cause of diarrhoea could only be established in 15–20 per cent of cases. Thanks to advances in laboratory techniques, an accurate diagnosis can now be made for at least 80 per cent of those who attend treatment centres which have access to appropriate laboratory facilities.

Different levels of investigation play a crucial role both in the treatment of individuals and in national control programmes. Knowing what organisms are causing diarrhoea makes surveillance easier. Epidemics and unusual infections can be spotted early and prompt warning given of the possible emergence of drug resistance among local strains of pathogens. Specific interventions can be more effectively planned, implemented and evaluated.

Good management

Success in planning and implementing diarrhoeal disease control programmes depends on good management at all levels. The World Health Organization is already providing senior management training courses for planners of national diarrhoeal disease control



Laboratory workers in Ethiopia

FAO photo

programmes. It is WHO's intention in the near future to offer appropriate management training to middle level health care personnel who will be responsible for the supervision of programmes in the field.

In this issue of *Diarrhoea Dialogue*, we look at some of the key points which health workers need to know about collection and transport of faecal specimens. The practical advice page describes simple, screening procedures which can be undertaken at any health centre possessing minimal equipment. At this level it is possible to make two important distinctions; between secretory and invasive diarrhoea (dysentery); and between bacillary and parasitic dysentery. These distinctions are very useful guides in decision-making about the immediate treatment of individual patients.

Targetting control measures

In all investigations, the closest collaboration must be ensured between clinical personnel, epidemiologists and microbiologists. Good epidemiological and clinical information speeds up and improves laboratory analysis of specimens. Similarly, the increased capability of laboratories to feed back an accurate diagnosis to the field permits more effective targetting of appropriate control measures. The complexity of the investigation that can be carried out in any particular situation obviously depends on the level of equipment and technical expertise that is available. A carefully planned and efficient network of laboratory services is thus a valuable component in any country's diarrhoeal disease control strategy.

K. M. E. and W. A. M. C.

In this issue . . .

- Basic laboratory techniques
- Reviews of laboratory manuals
- Health education and diarrhoeal diseases

AHRTAG

Appropriate Health Resources &
Technologies Action Group Ltd

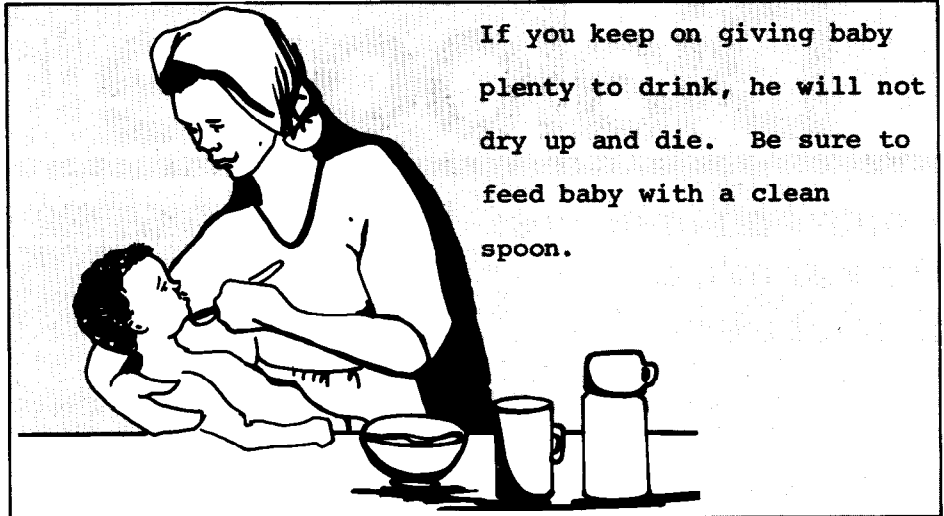
Success through education

The development, production and use of health education materials should be a major concern at all levels of diarrhoeal disease control programmes.* We have heard recently of various initiatives in this area.

● Jamaican materials

The Jamaican Ministry of Health and the Caribbean Food and Nutrition Institute have recently produced a set of teaching materials, funded by the WHO diarrhoeal disease control programme. The set contains:

- booklets with line drawings and photographs to tell mothers what to do when their children have diarrhoea and how they can recognize the signs and symptoms of dehydration.
- a manual for health workers containing technical information on diarrhoea and ideas on various ways that can be used to communicate messages to mothers.



An illustration from the Jamaican booklet.

If you keep on giving baby plenty to drink, he will not dry up and die. Be sure to feed baby with a clean spoon.

- a poster showing the signs and symptoms of dehydration to reinforce the message being put across by the health workers.

For more information about these materials contact: Elizabeth Grant, Field Co-Ordinator for Programme for the Control of Diarrhoea in Jamaica, Ministry of Health, 10 Caledonia Avenue, Kingston 5, Jamaica.

● Poster sets

Solange Muller has written to tell us about some poster sets being used in Nicaragua. The materials have also been tested in Mexico, the Dominican Republic and Honduras. The sets consist of two large posters (30cms × 40cms) one showing a thin dehydrated child and the other a healthy child, and at least ten small cards (15cms × 10cms) showing the causes of diarrhoea and ways of preventing it as related to conditions in that particular country. The small cards show simple line drawings of:

- a child defaecating outside
- a hand with dirty nails
- a baby bottle with flies on it
- a latrine
- food covered with a cloth
- a mother breastfeeding
- a mother giving oral rehydration solution.

If a large number of posters and cards are being made, they can be silk screened. But usually the health workers can make the materials themselves, by tracing from an original set and then colouring them in.

When using the posters, health workers should:

- show the mothers the poster of the thin dehydrated child
- ask for their comments
- ask whether there are any children in the community like this and why they get this way.

If mothers have problems in recognizing that the child is dehydrated because of diarrhoea, the health worker can give suggestions. The main idea is to

RUNNING BELLY CAN MAKE YOUR BABY DRY UP

WATCH FOR THESE SIGNS:—

IF YOU SEE ANY OF THEM, TAKE YOUR BABY AT ONCE TO THE NEAREST HEALTH CENTRE OR HOSPITAL

Showing mothers how to recognise dehydration.

encourage mothers to share ideas on diarrhoea.

Each mother then selects one of the small cards, explains what the drawing means to her and puts the card next to one of the two large posters, explaining why the card goes with the poster. After this, the mothers discuss with the health worker what they have learnt about preventing diarrhoea.

The set of posters and cards has been developed by Solange Muller and Felipe Orrego in the Dominican Republic for the Save the Children Fund, USA. For more information contact: Solange Muller, Apartado A-51, Managua, Nicaragua.

● Brazilian workshop

The Pan American Health Organization (PAHO) and the Centro Americano para Tecnología de Educación de Salud are organizing a workshop in Rio de Janeiro in December 1982. Participants will consider different ways of developing communications support for diarrhoeal disease programmes and produce some materials for field testing. There will be a report on the workshop in *Diarrhoea Dialogue 12*.

● WHO catalogue

The diarrhoeal disease control programme of WHO, Geneva is developing a catalogue of health education materials on the causes and prevention of diarrhoea. The catalogue shows posters, leaflets, journals and newsletters from all over the world. It will be available from WHO early in 1983. For more information contact: the Programme Manager, Diarrhoeal Disease Control Programme, World Health Organization, 1211 Geneva 27, Switzerland.

* Issue nine of *Diarrhoea Dialogue* gave a list of organizations who can supply information on health education materials.

In the next issue . . .

In 1883, research into cholera was made possible through Koch's discovery in Egypt of the comma bacillus or vibrio. Our first issue in 1983, *Diarrhoea Dialogue 12*, will review the achievements in cholera control over the past century. We will also look at parasitic diarrhoea.

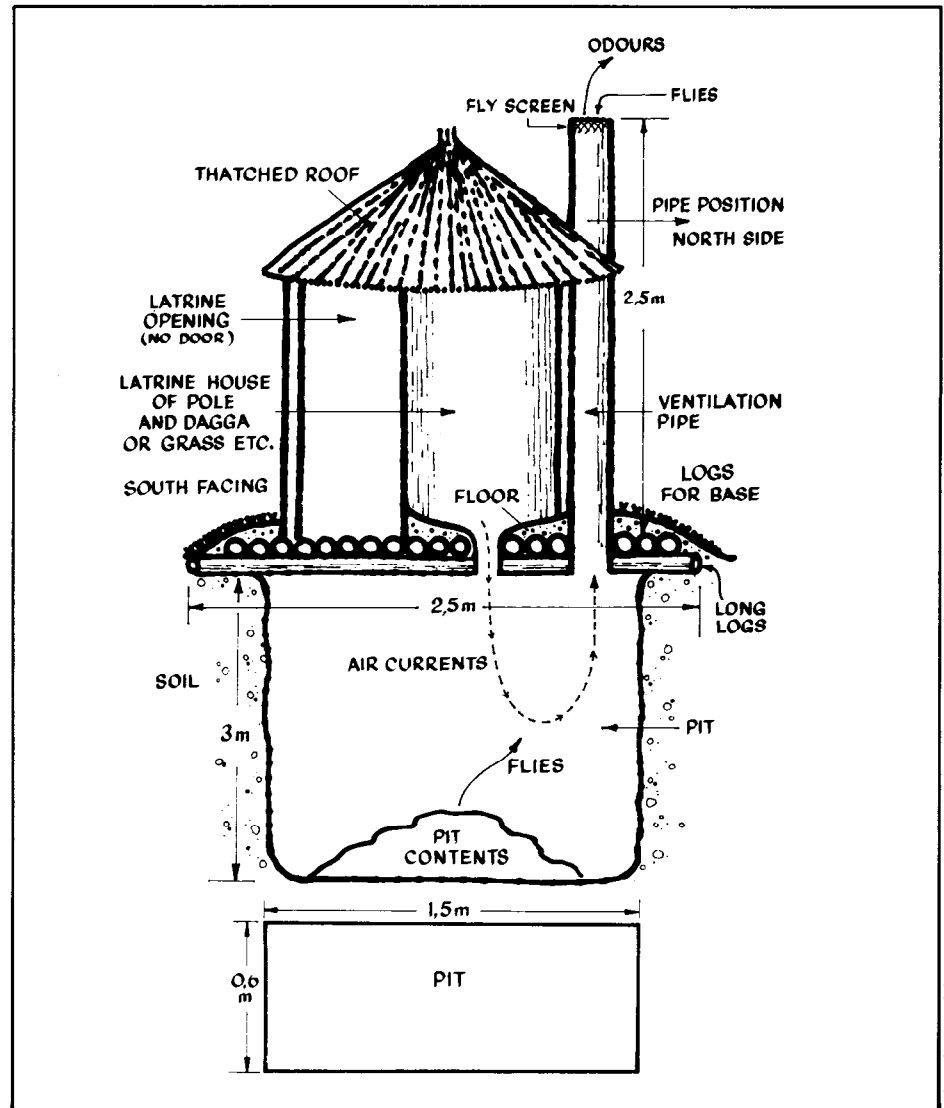
Zimbabwean latrines

Although pit latrine technology is basically simple, the effectiveness and acceptability of the designs that have been developed varies enormously.

One successful latrine has been produced in Zimbabwe by the Blair Research Laboratories. The superstructure is a doorless spiral with a large external vent pipe (see illustration). Lack of a door means that the latrine can always be kept dark—very important for fly control. Any flies that do manage to get into the pit escape via the external vent pipe. Ventilation is improved by the vent pipe and the absence of a cover (which can itself become contaminated) for the squat-hole.

More than 30,000 of these latrines have been built and are estimated to be in use in Zimbabwe. A superior permanent structure is available in kit form for around \$100.00, but the same design can also be built from mud and wattle or other local materials for around \$7.00. The building instructions issued by Blair Research Laboratories strongly emphasize how the system works to avoid mistakes in construction. If the latrine is built properly, and therefore works well, the likelihood of its being accepted and used is much greater.

For further information about the Zimbabwean latrine, write to Richard Middleton, Technical Advisory Group, World Bank, 1818 H Street, Washington DC 20433, USA.



The pit latrine developed by the Blair Research Laboratories in Zimbabwe

Collection, transport and examination

Laboratory services must work at all levels to be effective. Katherine Elliott and William Cutting consider some ways in which this can be achieved.

Laboratory services of varying complexity play their part in the investigation of diarrhoea at:

- primary health care centre level (especially useful in rural areas) using simple equipment to examine specimens and to prepare them for transport.
- intermediate referral level, usually at a district hospital's clinical laboratory with special equipment and materials for culturing bacteria and identifying them using a range of suitable tests.
- central level, where the public health reference laboratory uses sophisticated equipment and techniques (including electron microscopy) to complete the identification of the pathogen referred from clinical laboratories.

The value of an integrated approach

Most countries with diarrhoeal disease programmes will, in addition to their clinical laboratories, require a public health reference laboratory to investigate outbreaks of infectious diarrhoeal diseases, to assist with control and preventive measures, to supervise the testing of water supplies and to monitor and warn medical officers of the emergence of drug-resistant strains of major pathogens. As far as any national health budget permits, the central public health laboratory, or similar centre, should play a major part in investigating the epidemiology, treatment and prevention of diarrhoea-causing pathogens within the country.

If the problem of diarrhoeal disease control is approached in an integrated way from the outset, this will facilitate the most effective use of all available resources. This should include primary health care workers in remote, rural areas whose role — particularly in health education and the proper use of oral rehydration therapy — is crucial. Good communication is essential between all levels of any country's laboratory service to achieve and maintain reliability. Interchange of staff promotes better understanding of local constraints and helps to integrate the service as an effective whole.

Handling specimens

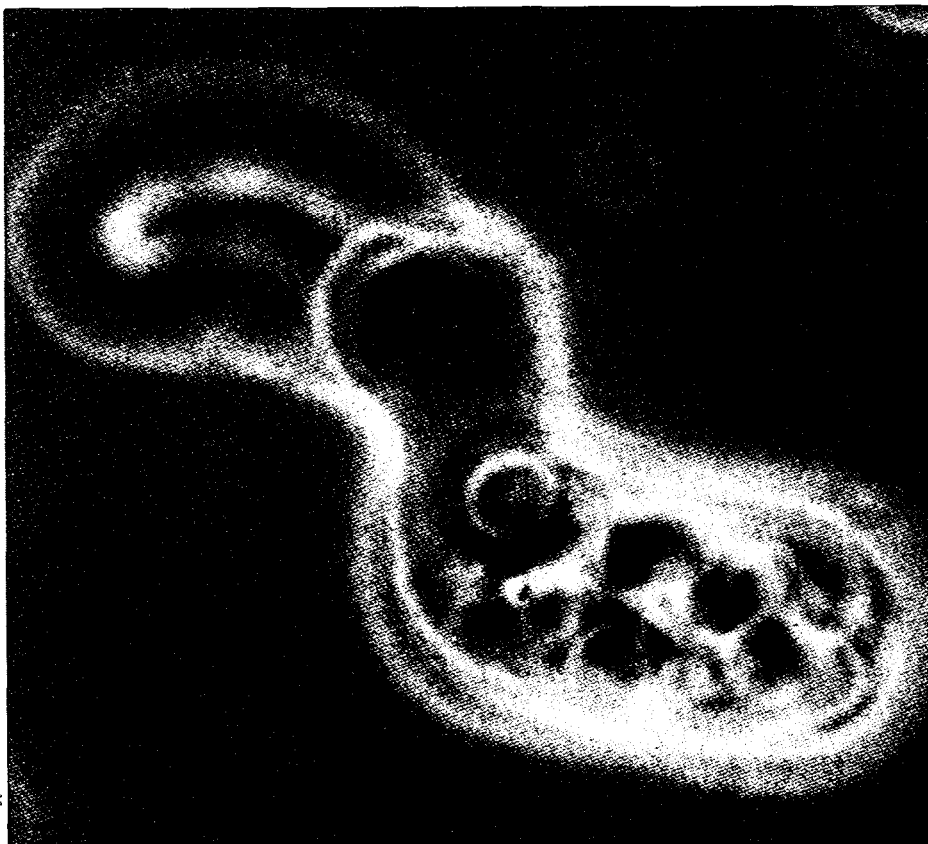
Health workers should know when and how to collect stool samples and how to send them quickly and safely to the nearest laboratory. The history of illness must be recorded accurately, indicating the presence or absence of fever, whether the problem is acute or chronic, and notes should be made about the appearance and volume of the freshly passed stool. It is possible to distinguish between dysentery (invasive diarrhoea with blood and pus in a small, soft or loose stool) and secretory diarrhoea which produces a very large liquid and watery stool.

At the health centre, simple equipment permits differentiation of bacillary dysentery, most often shigellosis, from dysentery due to parasites, especially *Entamoeba histolytica*. The practical advice page in this issue gives details of how to investigate diarrhoea at this level.

The time taken in transport is important. Specimens of faeces should reach the referral laboratory within two hours or otherwise be specially prepared for longer transport (see box). A health centre can prepare specimens in this way for transport if supplied with the necessary instructions and materials. The preparation of the appropriate media requires special facilities, including access to an autoclave and this process is normally undertaken at referral laboratories.

Laboratory technicians

The technician in any laboratory has a very responsible role. The results of tests play an important part in making decisions about treatment. Care, reliability and accuracy are essential.



Entamoeba histolytica amoeba containing ingested red cells (1 cm = approx. 5 microns)

Photo reproduced courtesy of Professor W. Peters, Professor H. M. Gilles and Wolff Medical Publishers Ltd.

Photo by Dr Stanley G. Browne.



Laboratory technicians play a very important role.

(Better to admit to a mistake in the laboratory than to run the risk of a mistake in treating the patient). Sound training is required and should be based on local health needs and the facilities that are likely to be available.

Safety must never be neglected in any laboratory, however simple. Equipment must be correctly installed and maintained. Specimens often contain highly infectious organisms and their safe disposal is of utmost importance. Laboratory workers must practise a high standard of personal hygiene which includes the washing of hands after handling specimens. Protective clothing should be worn and left in the

Collection

Specimens should be collected as soon as possible after the diarrhoea begins and before any antibiotics are given. A clean, dry and disinfectant-free receptacle must be used to collect the faeces. Contamination with urine should be avoided. The next step is to use a spoon, or wooden spatula, to transfer a portion of the stool, especially any part of it that contains blood or mucus, into a clean, dry, (need not be sterile) leak-proof container. Waxed paper cartons with lids, if available, are convenient but any leakproof small container will do. A piece of faeces about 15–20mm in diameter is enough — or, if the stool is liquid, about 5ml (in either case, a good teaspoonful). If worms, or segments of tape-worms can be seen, these should if possible be placed in a separate container. Containers must be clearly

Transport

If delay in transport to the appropriate laboratory cannot be avoided, two or more swabs from the stool should be inserted into a bijou bottle containing a suitable, sterile transport medium such as Stuart or Cary-Blair, breaking off the swab sticks to allow the bottle top to be tightly replaced. Cary-Blair medium preserves all enteric bacterial pathogens and prevents them from being overgrown by the normal bacteria always present in the bowel which do not cause diarrhoea. The media should always be inspected before use. If already cloudy, they are probably contaminated and useless. Try a different bijou bottle. Pathogens survive better in transport media if they can be kept as cool as possible and in the dark.

Salmonella, shigella, vibrio and yersinia organisms all survive well in Cary-Blair medium at ambient (normal surroundings) temperature for up to two days. Campylobacter survives for up to six hours only. If the specimen is for the culture of salmonella and shigella organisms only, buffered glycerol saline can be used as transport medium. If cholera is suspected, about 2ml of the fluid specimen should be

laboratory. Reagents and chemicals must not be mouth-pipetted and any eating, drinking or smoking in the laboratory should be forbidden.

labelled and be accompanied by a properly completed request form. The presence of blood, mucus, worms and tape-worm segments should be noted.

Where it is not possible to obtain a sample easily, for example from a baby, a specimen can be collected by inserting a cotton or alginate wool swab into the rectum for about ten seconds. Do not use lubricating gels. Make sure that the swab is well stained with faeces.

An alternative method is to insert a finger covered with a plastic fingerstall instead of a swab. Care should be taken to avoid unnecessary contamination of the specimen with bacteria from the skin surrounding the anus. The swab or fingerstall should then be placed in a container as above, breaking off the swab stick to fit.

placed into a container of sterile, alkaline peptone water which encourages the growth of vibrio organisms. If amoebic dysentery or other parasite infection seems likely and it is not possible to examine a fresh faecal specimen as described on page six, a small amount (0.5gm) of the stool should be transferred into a bijou bottle containing freshly prepared merthiolate iodine formaldehyde (MIF) fixative. This fixes protozoal parasites for later examination. Bacteria are also killed in MIF and faeces preserved in this medium cannot be cultured.

Where rotavirus is suspected about 1g of faeces (preferably from a stool rather than a swab) may be stored in phosphate buffered saline for analysis by the ELISA method.

Label clearly and pack properly in all instances, including always the matching, completed request forms — preferably in a polythene bag. Remember the bijou bottle contains live organisms! Great care should be taken to clean and disinfect everything that comes into contact with stool specimens from patients with diarrhoea. Faeces can be very dangerous and should be handled accordingly.

New techniques

The new API Z simple screening test of non-lactose fermenting colonies for salmonella and shigella is enzyme-based, commercially produced, gives a result after only two hours incubation and is relatively inexpensive.

A variety of new rotavirus screening tests for field use are now available as part of the ELISA range. The BIKEN Institute in Japan has developed a simplified test to identify enterotoxigenic *E. Coli* (ETEC).

If you would like more details about any of these tests, write to *Diarrhoea Dialogue*.

We would like to thank Bernard Rowe, Director, Central Public Health Laboratory, London; Peter Humphries, Principal MLSO at the Royal Free Hospital, London and Monica Cheesbrough, author of a *Medical Laboratory Laboratory Manual for Tropical Countries* for their advice and comments on this article.

Simple laboratory investigations into diarrhoea

Tony Moody looks at the range of laboratory work that can be carried out at health centre level.

The complete investigation of diarrhoea requires complex and sophisticated laboratory techniques to isolate and characterize the causative agent. This does not exclude the laboratory worker with only simple, basic equipment from being able to provide useful information for the management of diarrhoea.

Essential laboratory equipment consists of:—

- a microscope
- microscope slides and coverslips
- saline solution
- pipettes
- some basic stains and litmus paper

Total cost — less than \$500 US dollars.

Macroscopic appearance

Useful information can be gained by careful inspection of the faeces with the naked eye¹. The history of the patient's diarrhoea should be kept in mind whilst looking at the faeces. The *Clinician's Guide to Aetiology* published in *Diarrhoea Dialogue 7* and the new wall charts now available from the Ross Institute of Tropical Hygiene* may be useful aids in this preliminary screening.

- Profuse watery stools, sometimes flecked with mucus, occur in toxigenic *E. coli* diarrhoea and in cholera (rice water stool).
- Smaller, soft and frequent stools containing blood and mucus occur in amoebic dysentery and in bacillary dysentery due to shigella or campylobacter infections.
- Pale, frothy stools occur in giardiasis, tropical malabsorption and in lactase deficiency in infants.

Acid or alkaline reaction (pH) A simple litmus paper check on the stool pH can be helpful. Acid stools occur in amoebic dysentery and in lactase deficiency. In bacillary dysentery the stool is alkaline.

Microscopic examination

The stool should be as fresh and warm as possible when examined.

Saline preparation Place a drop of saline (0.9% sodium chloride) at room temperature onto a warm slide and, selecting an appropriate section of the faeces, i.e. from an area of bloody mucus or from the liquid part of the sample, use a stick to transfer a *small* amount to the slide. Emulsify this in the saline and place a coverslip over the preparation (discard the stick safely into disinfectant or burn). Examine the sample under the microscope using a 10x eyepiece and, initially, a 10x objective. Close the condenser iris diaphragm sufficiently to give good contrast. Careful searching of the whole coverslip area at this magnification could reveal active larvae of *Strongyloides*, ova of *Schistosoma* or other helminths, or clumps of pus cells and erythrocytes which indicate an inflammatory response to bacteria or amoebae.

Now turn the 40x objective into the viewing position and again search the whole coverslip area.

Dysentery At this level of investigation, the most important distinction that can be made is between amoebic and bacillary dysentery. Both stools may contain pus cells and macrophages and these show up well if a drop of 1% methylene blue stain is run under the coverslip. There may also be erythrocytes (red blood cells) in the stools.

In amoebic dysentery, if the specimen is fresh (still warm), very active trophozoites of *Entamoeba histolytica* (20–40 microns in size, i.e. about twice the size of polymorph leucocytes (pus cells)) ought to be easily observed moving rapidly across the slide, pushing out clear pseudopodia and containing ingested red blood cells. There should be no confusion between amoebae and macrophages which may also contain red blood cells. The amoebae move about and constantly change shape. The macrophages become immobile within a few seconds. In bacillary dysentery, caused by infec-

tions such as *Shigella*, *Campylobacter* and possibly invasive *E. coli*, there will be many pus cells, erythrocytes and macrophages, but no active amoebae or cysts. (Occasionally patients may have both amoebic and bacillary dysentery at the same time).

Flagellates Few or no cells in a fatty or unformed sample may indicate the cystic or trophozoite stages of *Giardia lamblia* in the saline preparation. The refractive axostyle and flagellar components of the cysts can be shown more clearly by using an iodine stain (1 in 5 dilution of Lugol's iodine in 10% acetic acid). A thin saline suspension of faeces smeared on a slide, air dried and fixed in methyl alcohol can be stained with a 1 in 20 dilution of Giemsa stain in pH 6.8 buffer for 15 minutes and will demonstrate the morphology of the trophozoites of *Giardia*. Fat is seen as yellow globules or fine needles and can be stained red with 1% alcoholic Sudan 111 stain.

Other flagellates may be present but are not pathogenic.

Vibrio cholerae Dark ground illumination and a hanging drop preparation are needed to identify the darting motility associated with cholera vibrios. A dark field condenser costs about £90 to modify a normal microscope. A temporary version can be contrived using plasticine or putty to use with the 40x objective (see page 61 in volume 1 of Monica Cheesbrough's *Medical Laboratory Manual for Tropical Countries* — reviewed on page seven).

Hanging drop preparation Take a slide and make a ring of about 1cm using vaseline. Place a coverslip flat on a table, add one drop of the liquid stool in the centre and place the vaseline ring on the slide over this drop. Quickly invert the slide and inspect under the 40x objective for the typical darting movement by rod-shaped organisms.

Tony Moody, Senior Chief MLSO, Hospital for Tropical Diseases, St Pancras Way, London NW1, U.K.

¹Banu et al 1982 *Epidemiologic and clinical features of patients infected with shigella who attended a diarrhoeal diseases hospital in Bangladesh. The Journal of Infectious Diseases, Vol. 146 no 2: 177–183.*

**Inquiries to Dr Isabelle de Zoysa, Ross Institute of Tropical Hygiene, London School of Hygiene and Tropical Medicine, Keppel St, London WC1, U.K.*

Books for medical laboratory workers

Peter Humphries reviews three laboratory manuals that include investigation techniques for diarrhoea.

Medical Laboratory Manual for Tropical Countries (Volume 1, first edition 1981. Volume 2 in preparation — due for publication in June 1983)
Monica Cheesbrough (author and publisher)

Price: £4.70 (developing countries)*
 £7.60 (other countries)*

The author has kindly provided pre-publication text for review from the microbiology section of Volume 2.

Volume 1 contains the parasitology of stools and fully describes the helminths and protozoa causing diarrhoea. Methods are carefully and singly explained, well illustrated with detailed notes on likely problems. The author's personal experience of most of the techniques given is clearly demonstrated. The text is up-to-date and presented in a friendly style. Safety procedures are a welcome inclusion, too often omitted from descriptive manuals. In addition to its full technical repertoire the text includes introductory material and explains the clinical relevance of the laboratory procedures described.

The microbiology section previewed from Volume 2 includes a definitive investigation of diarrhoea as a clinical entity. Modern ideas about tropical diarrhoea are fully explained together with associated laboratory procedures which include instructions for workers at all levels of health care. Technical matters are absolutely up-to-date and will prove useful for the modern laboratory anywhere in the world. Again, the author has achieved a friendly, communicative style without losing the authority which underlies all the material in the section reviewed.

Of the three manuals reviewed, these two volumes by Monica Cheesbrough can be recommended for medical laboratory workers anywhere.

Whatever facilities they have, there will be something to improve their services to health care.

*Postage and packing for developing countries is £1.25 and £1.75 elsewhere. You can obtain the manual by writing to *Monica Cheesbrough, 14 Bevills Close, Doddington, Cambridgeshire PE15 0TT, UK. Visitors to London can obtain the manual from the Institute of Child Health, 30 Guildford Street, London WC2.*

Manual of Basic Techniques for a Health Laboratory 1980
(New edition in preparation)
World Health Organization
 Price: 30 Swiss Francs (approximately £8.50)*

This book is a mixture of reference material and illustrated techniques, both aspects freely intermingled, a feature which is sometimes confusing.

Diarrhoea is not treated as an entity and does not appear in the index, but it does appear in various places when the condition is associated with the parasite under discussion.

The reference aspect is extremely detailed with useful size comparison charts for helminth ova and pictures of artefacts. Style is strictly factual which can be expected in a laboratory reference manual. Technical instructions are also described in an impersonal style, leaving out much important detail, in contrast to the reference text.

Sections on concentration methods are well illustrated and given for a variety of infections including strongyloides larvae. Excellent instructions are given for despatch of stools for parasites and bacterial cultures to laboratories where these investigations can be carried out.

There are some surprising omissions

for such a recent text, particularly on clinical links with the investigations described. This is a book for the laboratory shelf, rather than a friendly companion.

* You can obtain this manual by contacting your WHO regional office or writing directly to WHO headquarters in Geneva. Postage will depend on where the book is being sent. WHO, 1211 Geneva 27, Switzerland.

A Medical Laboratory for Developing Countries 1972 (latest edition)

Maurice King
Oxford University Press
 Price: £3.50*

The word diarrhoea does not appear in the index, nor is the condition treated as an entity. However, Chapter 10 deals fully with the examination of stools, concentrating on macroscopic and microscopic methods for helminths and protozoa. The text is very detailed, accompanied by easily followed diagrams and pictorial sequences of techniques. The formaldehyde-ether method is the only concentration technique described and its limitations are not fully explained.

The coverage of diarrhoea is limited to bacillary and amoebic dysentery, giardiasis and lactase deficiency in infants. The importance of macroscopic appearance of stools for signs of helminths or exudates is emphasized and a method for estimating the pH of stools is described. Stool culture for bacteria is omitted and no instructions are given for stool transport to laboratories where this can be done.

There are many tips for carrying out the procedures described with homemade tools and the style of the text is simple and friendly. Nevertheless, conceptual and technical advances in the investigation of tropical diarrhoea have overtaken some of the fundamental ideas in this book, due mainly to the length of time since its publication.

* Postage will depend on where the book is being sent. For details write to *Oxford University Press, 37 Dover Street, London W1, UK.*

Peter Humphries, Principal MLSO, Royal Free Hospital, Pond Street, London NW3, UK.

ORT in Tamil Nadu

We very much appreciate oral rehydration therapy in diarrhoeal illnesses. This is the first measure we have taken in primary health care along with our leprosy control work. We advise the mothers to use a *palade* (a small metal vessel used for feeding young babies) to measure sugar and common salt and an empty gripe water bottle to measure out adequate water. Both the *palade* and the bottle are common household articles in Tamil Nadu. These are the quantities we use to prepare the oral rehydration drink:

- one level *palade* full of sugar (7.0gm)
- one level *palade* spout-full of salt (1.0gm)
- one small pinch of baking soda
- dissolved in one gripe water bottle of clean water (130ml)

This drink is very palatable and well accepted by children. We are very happy to share this information with you.

Dr Mohan, Kasturba Nilayam, Malavanthangal, Via Kandachipuram, S.A.Dt. 605701, T.N., India.

Village programmes

We have nine expert nurse midwives here and an auxiliary training programme with 15 students at present. Mass Media from the USA are distributing 200,000 pictures showing how to make sugar-salt solution. In many villages the person who has been shown how to make up the solution has a flag (a black baby on a pink background) so that people in the compound know where to go for help. We find that, after instruction, mothers are giving the sugar-salt solution, but sometimes continue it for longer than recommended before getting further help.

Instructions for making up the solution have also been given on Radio Gambia and there is certainly a great deal of interest.

Dr John E. Matthias, Banjul, The Gambia, West Africa.

Soya milk

I was surprised that no mention was made on page six of *Diarrhoea Dialogue 6* (*Persuading children with*

diarrhoea to eat) of soya beans. We find them a real godsend, especially for making soya milk to give to children with diarrhoea. This 'milk' is high in protein content and lactose-free. It contains about three quarters of the calcium supplied in breast milk and also has a satisfactory iron and Vitamin B content.

We cannot unreservedly recommend soya milk as the sole food for infants under four months, who, in any case should be breastfed. However, it is extremely useful for alternative or temporary feeding, or for infants with diarrhoea. We have included a recipe for making soya milk so that other people can try it.

1. Wash, sort and soak 500gm soya beans in plenty of clean water overnight.
2. Mince or grind the beans.
3. Boil two litres of water and add the minced beans. Bring back to the boil. Stir and be careful the mixture does not boil over. Boil for 15 minutes.
4. Strain through a clean cloth or wire sieve. Add a pinch of salt and some sugar to taste.

Anna Pearce, Compassion, Brooklyn, Capetown, South Africa.

Traditional remedy in India

An article in DD 6 about the use of rice water and rice porridge for rehydration reminded me of my grandmother's remedy, which saw three generations of children safely through diarrhoea episodes, usually without any other medication. This is a porridge of sago (known locally as saboodana or sabbiyam) cooked in water to a gelatine-like consistency, served with salt and sugar to make it palatable. Milk or buttermilk were added as soon as the condition improved, to take care of the need for protein. 4-5 helpings were given, 8oz for adults and proportionately less for children. Most children enjoy the bland flavour of this preparation, it is highly effective and cheap.

Ms Srilatha Batliwala, Bombay 400 018, India.



Health workers being trained in Tamil Nadu

Editors: Dr William Cutting (U.K.) and Dr Katherine Elliott (U.K.)

Editorial advisers: Dr David Candy (U.K.), Dr I. Dogramaci (Turkey), Dr Richard Feachem (U.K.), Dr Michael Gracey (Australia), Dr N. Hirschhorn (U.S.A.), Dr D. Mahalanabis (India), Dr Leonardo Mata (Costa Rica), Dr Mujibur Rahaman (Bangladesh), Dr Jon Rohde (Rockefeller Foundation), Ms. E. O. Sullesta (Philippines), Dr Paul Vesin (France), Dr M. K. Were (Kenya).

Executive editor: Denise Ayres With support from WHO, UNDP, GTZ and SIDA

Diarrhoea Dialogue 

Diarrhoea Dialogue is produced by AHRTAG at 85 Marylebone High Street, London W1M 3DE. Tel. 01-486 4175.